Synthesis of Stereoselectively Labelled Citric Acids transformable into Chiral Acetic Acids

Svante Brandänge* and Olof Dahlman

Department of Organic Chemistry, Arrhenius Laboratory, University of Stockholm, S-106 91 Stockholm, Sweden

The ketones (1b) and (1c) have been transformed into the monodeuteriated citric acids, (4b) and (4c) respectively, which are suitable starting materials for the synthesis of chiral acetic acids and other labelled compounds; under base catalysis, the axial 2-H in (1a) exchanges with deuterium more than 1000 times faster than the axial 2-H in 4-t-butylcyclohexanone.

The use of stereoselectively isotope-labelled compounds is a powerful technique for the elucidation of various stereochemical phenomena. We report here that a synthetic route (Scheme 1) which has previously been used for the synthesis of citric acid and some of its analogues¹ is also suitable for the preparation of stereoselectively monodeuteriated citric acids (4b) and (4c) and the enantiomer of (4c). In addition, we report an unusually fast base-catalysed exchange of the axial 2-H in (1a) with deuterium.

When the stereoselectively monodeuteriated ketone (1b)² was transformed according to the reaction sequence (1) \rightarrow (2) \rightarrow (4) in Scheme 1, we obtained (1S,2S)-[1-²H₁]-2-hydroxypropane-1,2,3-tricarboxylic acid (4b) { $[\alpha]_{578}^{20} - 0.4^{\circ}$, $[\alpha]_{436}^{20} = -0.77^{\circ}$ (c 2.5, H₂O); in saturated aq. (NH₄)₂MoO₄¹: $[\alpha]_{436}^{20} = -3.11^{\circ}$ (c 2.5)}. The ¹H n.m.r. spectrum (Figure 1) corroborates the high stereochemical purity of (4b) and shows in addition that the δ 3.04 and 2.86 signals of the AB spectrum must be due to H_a and H_b, respectively; the assignment for citric acid (4a) is obtained by replacing D with Ha and H with H_b in (4b). The introduction of a deuterium atom into citric acid leads to a small (0.018 p.p.m.) upfield shift of the signal from the geminal proton. The trimethyl ester (CH₂N₂) of (4b) was obtained in a 20% overall yield from the epoxide precursor of (1b) and was contaminated with 4-5% of the nondeuteriated analogue (mass spectra).

Degradation of the minor Reformatsky product (3b) obtained from (1b) (Scheme 1) gave the enantiomer of (4c), contaminated with 3-4% of the nondeuteriated analogue (mass spectra) {found: $[\alpha]_{578}^{20} + 1.1^{\circ}, [\alpha]_{436}^{20} + 1.6^{\circ} (c \ 2.5, H_2O);$ in saturated aq. $(NH_4)_2MOO_4^{-1}: [\alpha]_{436}^{20} + 29.4^{\circ} (c \ 2.5)$ }. A 65% deuteriated sample of this citric acid was synthesised previ-

ously by axial deuteriation and subsequent oxidation of 3-dehydroquinic acid.³ A similar sequence involving dideuteriation-monodedeuteriation gave the enantiomer of (4b) which was 70% deuteriated.³ These samples were not characterised further.

Compound (1b) can also be obtained in a less pure form $(4.9\% [^{2}H_{0}], 87.3\% [^{2}H_{1}], 7.8\% [^{2}H_{2}])$, by treatment (10 min) of (1a) (200 mg) with potassium carbonate (61 mg) in D₂O-CH₃OD-tetrahydrofuran (2:2:10 ml) at 10 °C. By analogy with results from other six-membered ring ketones,⁴ there was a rapid exchange of the axial 2-H and a slower exchange of the equatorial 2-H (¹H n.m.r. spectroscopy). A rate ratio $k_{\rm ax}/k_{\rm eq}$ of about 35⁺ was found by mass spectral analysis of the mixture of deuteriated trimethyl citrates obtained by degradation of the mixture of deuteriated ketones (1). This ratio is smaller than those obtained for ketones which are presumably conformationally less flexible.⁴ However, the reactivity of the axial 2-H is unusually high. Ketone (1a) (50 mg) or 4-t-butyl-cyclohexanone (30 mg) in dioxane (5 ml) was treated at 11.5 \pm 0.5 $^\circ C$ with 1.25 ml of a solution of NaOD in D_2O which showed 11.2 on the pH meter. Aliquots were acidified to ca. pH 6 and worked up and the formation of (1b) or deuteriated 4-t-butylcyclohexanone was studied by ¹H n.m.r. or mass spectroscopy, respectively. The half-lives found for (1a) and 4-t-butylcyclohexanone were of the order of 1 min and 30 h, respectively. A study of the cause of this difference in reactivity is in progress.

Isomer (4c) was prepared from (1c), which was obtained

[†] This value is approximate only, the main reason being the unexpected consumption of base during the exchange reaction.



a; $R^1 = R^2 = H$. **b**; $R^1 = D$, $R^2 = H$. **c**; $R^1 = H$, $R^2 = D$.

Scheme 1. Synthesis of the citric acids (4) from the ketones (1). Reagents: i, Zn, BrCH₂CO₂Et; ii, HOAc, H₂O, heat; iii, KMnO₄, aq. NaOH (ref. 1). The Reformatsky reaction produces (2) and its C-3 epimer (3) in the ratio 94:6 (ref. 1). The overall yield (1) \rightarrow (2) \rightarrow (4) is about 25% (ref. 1).



Figure 1. Methylene region of a 200 MHz 1 H n.m.r. spectrum of the monodeuteriated citric acid (4b) in D₂O.

by dideuteriation¹ of (1a) followed by treatment with potassium carbonate as for (1b) but for 36 min and in nondeuteriated solvents. As found by mass spectral analysis of the trimethyl ester (CH₂N₂), the sample of (4c) had the following composition: 17.9% [²H₀], 81.2% [²H₁], 0.9% [²H₂] {found for this mixture: $[\alpha]_{578}^{20} - 0.5^{\circ}$, $[\alpha]_{436}^{20} - 1.0^{\circ}$ (*c* 2.6, H₂O); in saturated aq. (NH₄)₂MOO₄⁻¹: $[\alpha]_{436}^{20} - 20.8^{\circ}$ }.

The technique presented here is the first to allow stereoselective introduction of a deuterium atom into the *pro-S* carboxymethyl group of citric acid. The new compounds



Scheme 2. Enzymatic conversions of (4b) into (R)- $[2-{}^{2}H_{1},{}^{3}H_{1}]$ -acetic acid or (4S)- $[4-{}^{2}H_{1}]$ -L-glutamic acid.

(4b) and (4c) can be transformed into chiral acetic acids⁵ by using the enzyme citrate lyase (commercially available) in tritiated water (Scheme 2). To determine the inversion stereochemistry of this enzymatic reaction, tritiated analogues of (4b) and (4c) were prepared by means of a three-⁶ or fourenzyme⁷ synthesis which included the use of the enzymes aconitate isomerase⁶ or *re*-citrate synthase⁷ which are difficult to obtain.

It should also be possible, by means of enzymatic conversions of (4b) or (4c), to prepare stereoselectively deuteriated samples of aconitic acid, isocitric acid, α -ketoglutaric acid, and glutamic acid (Scheme 2) (cf. ref. 8).

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References

- 1 S. Brandänge, O. Dahlman, and L. Mörch, J. Am. Chem. Soc., 1981, 103, 4452.
- 2 R. F. Butterworth, P. M. Collins, and W. G. Overend, J. Chem. Soc., Chem. Commun., 1969, 378.
- 3 E. Haslam, M. J. Turner, D. Sargent, and R. S. Thompson, J. Chem. Soc. C, 1971, 1489.
- 4 G. Lamaty, 'Isotopes in Organic Chemistry,' vol. 2, eds. E. Buncel and C. C. Lee, Elsevier, Amsterdam 1976, p. 33; J. Toullec, Adv. Phys. Org. Chem., 1982, 18, 1.
- 5 H. G. Floss and M.-D. Tsai, Adv. Enzymol. Relat. Areas Mol. Biol., 1979, 50, 243; H. G. Floss, Methods Enzymol., 1982, 87, 126; K. Kakinuma, N. Imamura, and Y. Saba, Tetrahedron Lett., 1982, 1697; P. F. Leadlay and S. Greer, Anal. Biochem., 1982, 127, 89; J. D. Rozzell, Jr., and S. A. Benner, J. Org. Chem., 1983, 48, 1190.
- 6 J. P. Klinman and I. A. Rose, Biochemistry, 1971, 10, 2267.
- 7 W. Buckel, H. Lenz, P. Wunderwald, V. Buschmeier, H. Eggerer, and G. Gottschalk, Eur. J. Biochem., 1971, 24, 201.
- 8 E. Ziegler, H. Gubbels, and H. J. Reisener, Z. Pflanzenphysiol., 1976, 77, 278.